

Disclosing molecular alterations in the blood-brain barrier endothelium during interaction with breast cancer cells: a step forward brain metastasis prevention

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Background

Brain metastases are amongst the impact factors associated with decreased life expectancy of many breast cancer (BC) patients. The extravasation, a key step for the establishment of metastasis, is thought to involve the disruption of the microvascular endothelium that composes the blood-brain barrier (BBB) by cancer cells. However, little is known about the mechanism involved in BBB transposition by BC cells.

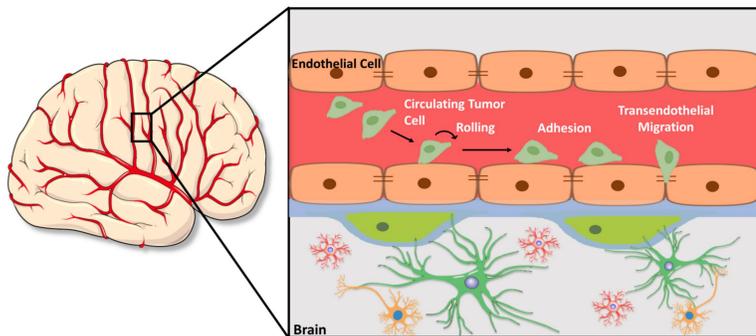
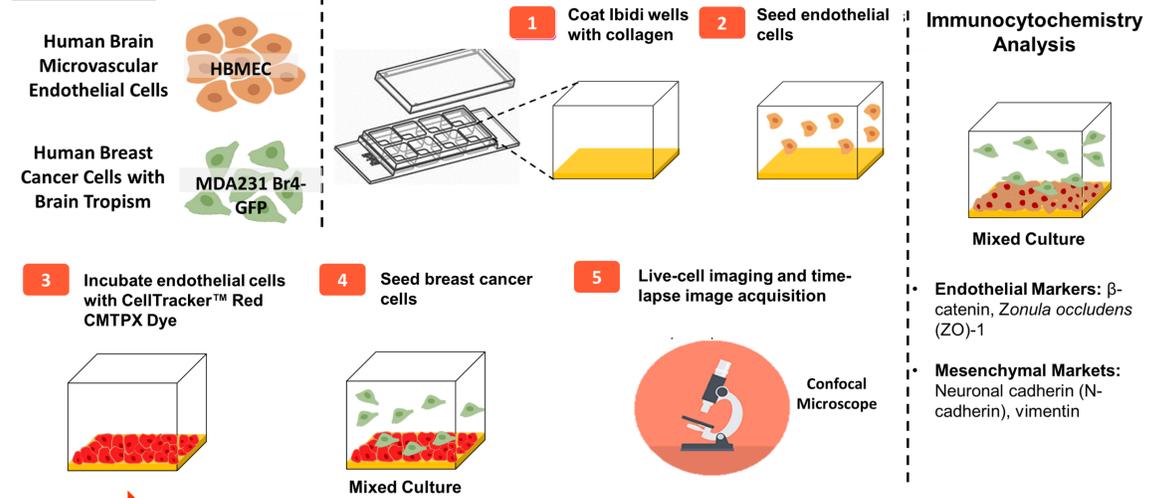


Fig. 1 | Schematic representation of cancer cells extravasation across blood-brain barrier (BBB), a multi-step process involved in brain metastases development.

Methods



Aim

Dissect the interaction between BC cells and BBB endothelial cells

Results

BC cells acquired a highly invasive phenotype during interaction with BBB endothelial cells

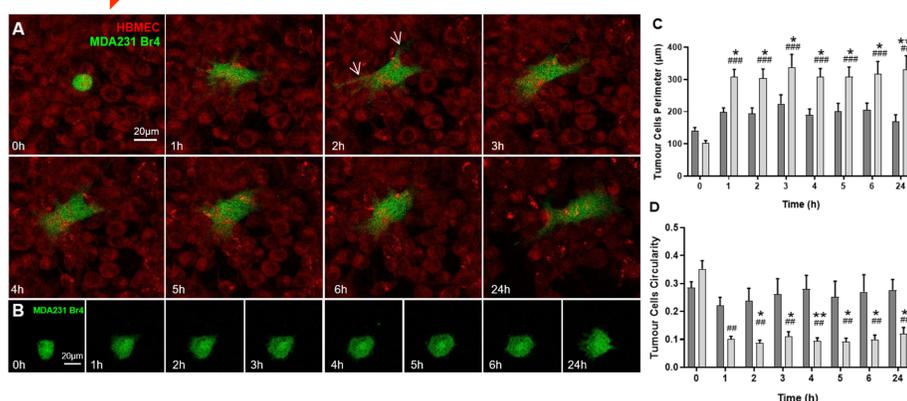


Fig. 2 | MDA231 Br4 cells acquire an invasive phenotype during extravasation. Temporal profile of interaction between MDA231 Br4 cells (tagged with GFP; green) and HBMEC confluent monolayers (labelled with CellTracker™ Red CMTPIX Dye; red) in mixed (A) and single cultures of MDA231 Br4 cells (control) (B) at 0, 1, 2, 3, 4, 5, 6 e 24 h. Morphological parameters such as tumour cells perimeter (C) and circularity (D) were analysed at the same timepoints. Arrows indicate the cytoplasmic cell extensions. Statistical significances are denoted as *P<0.05, **P<0.01 and ***P<0.001 for mixed culture vs control at the same timepoint and #P<0.05, ##P<0.01 and ###P<0.001 for mixed culture of indicated timepoints vs. mixed culture at 0 h.

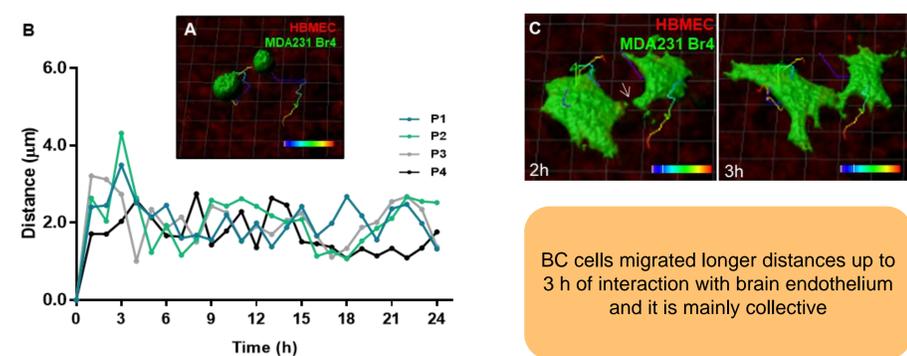


Fig. 3 | MDA231 Br4 cells acquire a migratory phenotype during extravasation. 3D representation of the total distance migrated by MDA231 Br4 cells (tagged with GFP; green) along 24 h (A). Semi-quantitative analysis of MDA231 Br4 cells migrated distances along 24 h (B). Each line in B represents the distances mean of four MDA231 Br4 cells in four different positions on ibidi wells. Representation of tumour cells collective migration at 2 and 3 h of interaction (C). Arrows indicate the contact points between tumour cells.

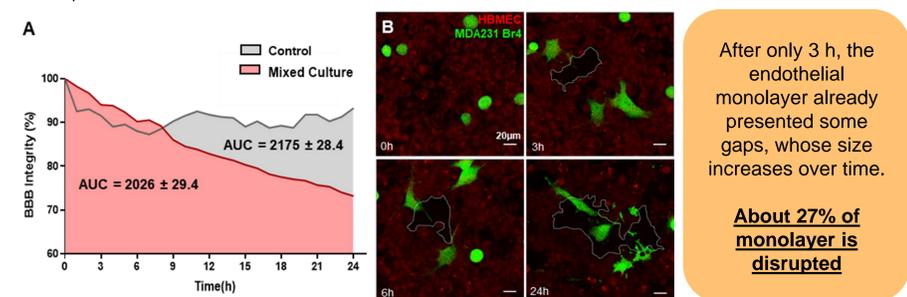


Fig. 4 | HBMEC and MDA231 Br4 cells interaction decrease the blood-brain barrier (BBB) integrity. The effect of malignant-endothelial cells interaction on BBB integrity was evaluated in mixed and endothelial cultures (control) by live-cell imaging microscopy and area under curve was determined along 24 h (A). Tumor cells transmigration promoted an impairment of BBB along time, confirmed by the increase of monolayer holes size upon the interaction with MDA231 Br4 cells (circled with a white line) (B).

Conclusion

These findings indicate that along extravasation BCCs acquire invasive properties, while the endothelium undergoes profound phenotypic changes, which altogether impact on BBB integrity and culminate in BCCs transmigration.

BC cells localize at different endothelium levels during the extravasation process

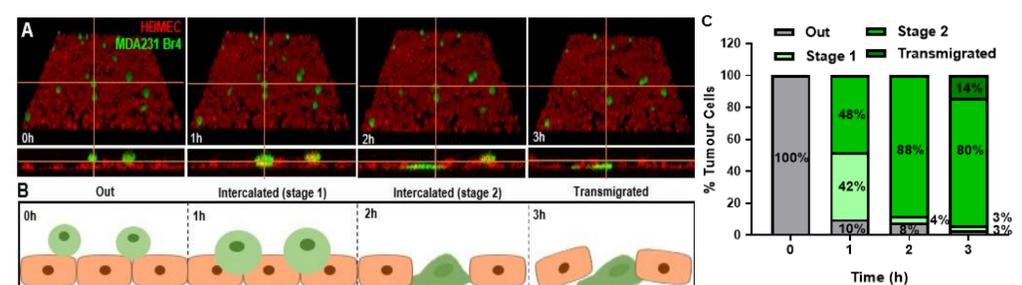


Fig. 5 | MDA231 Br4 cells localize at different endothelium levels during extravasation. 3D and orthogonal visualization of MDA231 Br4 cells (tagged with GFP; green) interaction with HBMEC confluent monolayer (labelled with CellTracker™ Red CMTPIX Dye; red) at 0, 1, 2 and 3 h (A). Schematic representation of tumour cells position relative to confluent HBMEC cultures (out, intercalated stage 1, intercalated stage 2 and transmigrated, corresponding the tumour cells over, partially or completely inserted in the endothelial monolayer, below the endothelium) (B). Semi-quantitative analysis of tumour cells positions relative to endothelium (out and intercalated stage 1, stage 2 and transmigrated) (C) at the same timepoints.

BBB endothelial cells undergone a Endothelial-Mesenchymal Transition that favors the paracellular transmigration

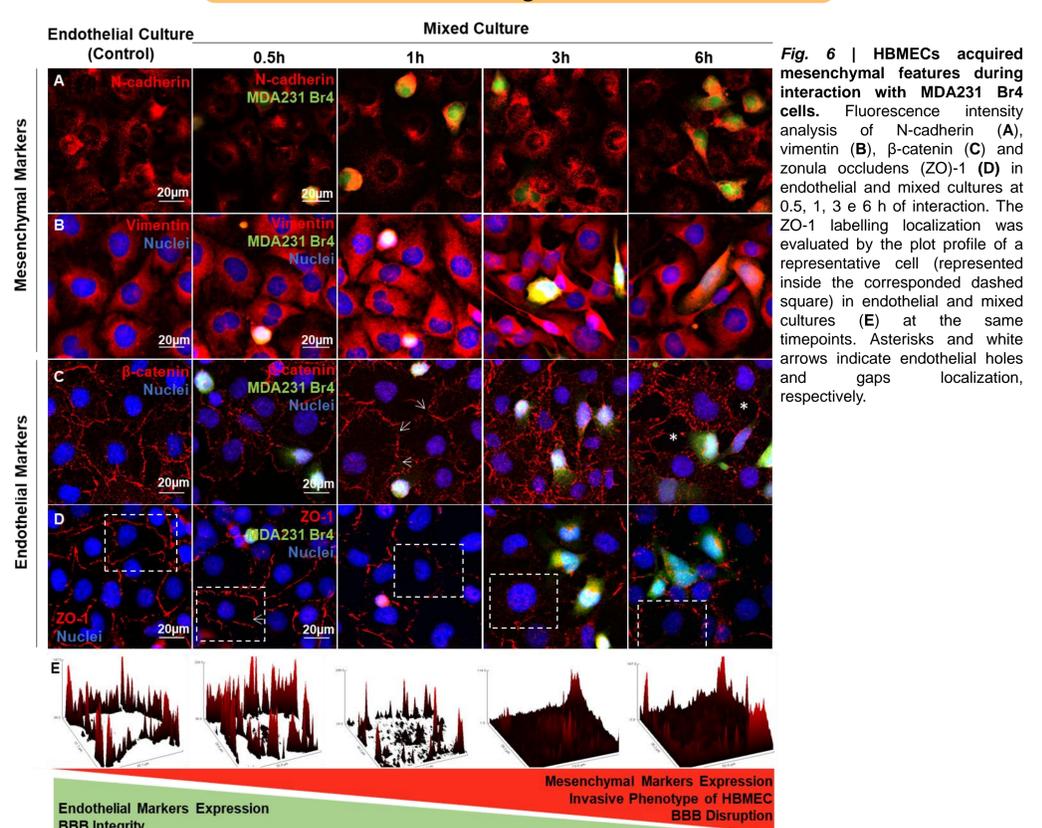


Fig. 6 | HBMECs acquired mesenchymal features during interaction with MDA231 Br4 cells. Fluorescence intensity analysis of N-cadherin (A), vimentin (B), β-catenin (C) and zonula occludens (ZO)-1 (D) in endothelial and mixed cultures at 0.5, 1, 3 e 6 h of interaction. The ZO-1 labelling localization was evaluated by the plot profile of a representative cell (represented inside the corresponded dashed square) in endothelial and mixed cultures (E) at the same timepoints. Asterisks and white arrows indicate endothelial holes and gaps localization, respectively.