

Fundação

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Medicinal plants from the Guinea-Bissauan flora: Anti-inflammatory properties of Parinari excelsa and the chemistry behind it

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INTRODUCTION

encontro

CIENCIA

METHODS

Parinari excelsa stem bark hydroethanol (1:1) extract preparation

Guinea plum, scientifically named Parinari excelsa, is a tree from the Chrysobalanaceae family that can reach 25 m [1, 2]. It is widespread across West Africa, where it is employed for wound healing and in the treatment of several health conditions, such as chest pain, malaria and diabetes [1,3,4]. This plant is one of the Guinean-Bissauan species studied by our research group. A hydroethanol extract from its stem bark caught our attention due to its score in an in vitro screening of antiinflammatory activity performed with extracts from several species from Guinea-Bissau. In this work we investigated its effects on the expression of inflammatory mediators and we studied its chemical composition.

1. In vitro cell-based assays	2. Chemical characterization and compound isolation
	24 10 0
1.1. ELISA for the evaluation of the	2.1. HPLC
expression of the inflammatory mediators	2.2. MS
IL-6 and TNF- α in THP-1 cells;	2.3. TLC
1.2. Luminescence-based assay for the	2.4. NMR:
evaluation of the NF-кВ activity	2.4.1. 1H NMR
(macrophages)	2.4.2. 1C NMR
	2.4.3. DEPT 135
	2.4.4. COSY
	2.4.5. HSQC
	2.4.6. HMBC

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RESULTS

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С

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Figure 1. Effects of the hydroethanol extract of *P. excelsa* on the expression of IL-6 and TNF-α in THP-1 derived macrophages. IL-6 and TNF-α results are expressed as mean ± standard error of three independent experiments, performed in duplicate. ***p<0.001 compared to the LPS stimulated control.



Concentration (µg/mL)

Figure 2. Effects of the hydroethanol extract of P. excelsa on the NF-KB activity in THP-1 Lucia derived ults are expressed as mean ± standard error of three independent experiments, performed macrophages. Results are expressed as mean ± standard error in triplicate. ***p<0.001 compared to the LPS stimulated control.

CONCLUSIONS AND FUTURE PERSPECTIVES

The extract from the stem bark of P. excelsa inhibited the NF-kB pathway and reduced the levels of IL-6 and TNF-α.

- In the HPLC-DAD chromatogram a detached and major compound was noticed, with UV spectrum characteristic of a flavonoid.
- Naringenin-8-sulfonate was isolated for the first time from a species of Parinari genus.
- Our next goal will be to access the anti-inflammatory ability of the isolated compound to understand if it may be, at least partially, responsible for the anti-inflammatory activity of the extract.

Time Compound 1: C₁₅H₁₂O₈S D 0=





Figure 3. Chemical characterization and compound isolation. A - HPLC-DAD chromatogram (280 nm) of the hydroethanol extract obtained from the steam bark of *P. excelsa*. B - *P. excelsa* extract's TLC at 365 nm. C - HPLC-DAD chromatogram (280 nm) of the compound obtained after TLC purification. D - Chemical structure of the isolated compound, identified as naringenin-8-sulfonate.

REFERENCES

[1] Ndiaye, M., Diatta, W., Sy, A.N., Dièye, A.M., Faye, B., Bassène, E., 2008. Antidiabetic (1) товаув, т., отака, т., оу, к.т., отвус, к.т., гауе, Б., Bassene, Е., 2008. Antidiat properties of aqueous barks extract of *Parinari excels* in alloxan-induced diabetic rats. Fitoterapia. 79, 267-270.

[2] Wyk, B. E., 2015. A review of commercially important African medicinal plants. J Ethnopharmacol. 176, 118-134.

[3] Boer, H. J., Kool, A., Broberg, A., Mziray, W. R., Hedberg, I., Levenfors, J. J., 2005, Antifungal and anti-bacterial activity of some herbal remedies from Tanzania. J Ethnopharmacol. 96, 461-469.

[4] Kamuhabwa, A., Nshimo, C., Witte, P., 2000. Cytotoxicity of some medicinal plant extracts used in Tanzanian traditional medicine. J Ethnopharmacol. 70, 143-149.

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